

Synthesis and utility of fluorogenic acetoxyethyl ethers

Luke D. Lavis, Tzu-Yuan Chao, and Ronald T. Raines*^a

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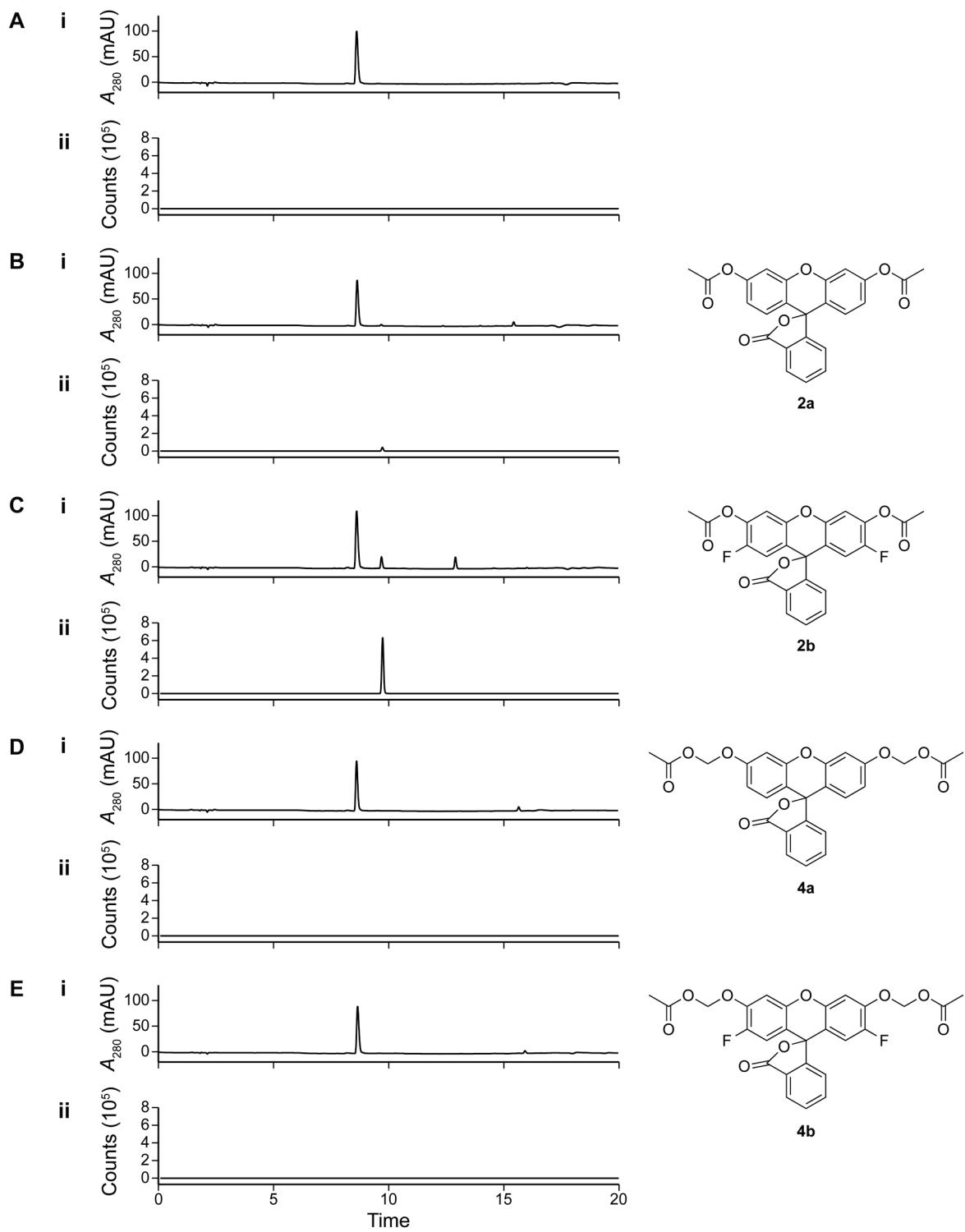


Fig. S1 Chemical reactivity of acetate esters **2a** and **2b** (100 μ M), and AM ethers **4a** and **4b** (100 μ M) with Ac-Arg-Phe-Met-Trp-Met-Lys-NH₂ (1 mg/mL) in 10 mM HEPES buffer, pH 7.3, for 2 h. (i) HPLC chromatogram of absorbance at 280 nm. (ii) LC-MS chromatogram with single ion monitoring-mass spectrometry (SIM-MS) for acylated peptide ($m/z = 981.47$). (A) Peptide only. (B) Acetate ester **2a**. (C) Acetate ester **2b**. (D) AM ether **4a**. (E) AM ether **4b**.

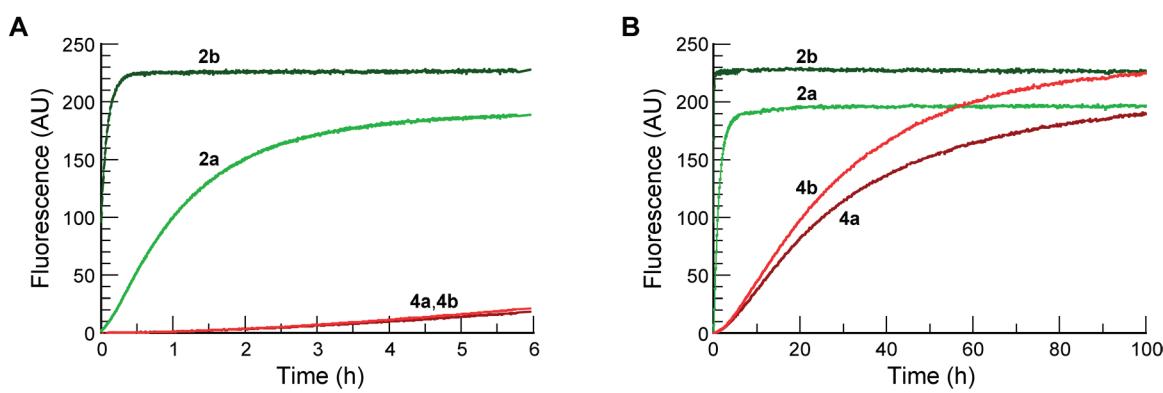


Fig. S2 Time course for the spontaneous generation of fluorescence (λ_{ex} 490 nm, λ_{em} 520 nm) from acetate esters **2a** and **2b** (25 nM), and AM ethers **4a** and **4b** (25 nM) in DMEM containing 10% v/v FBS. (A) 0–6 h; this panel replicates Figure 1B. (B) 0–100 h.

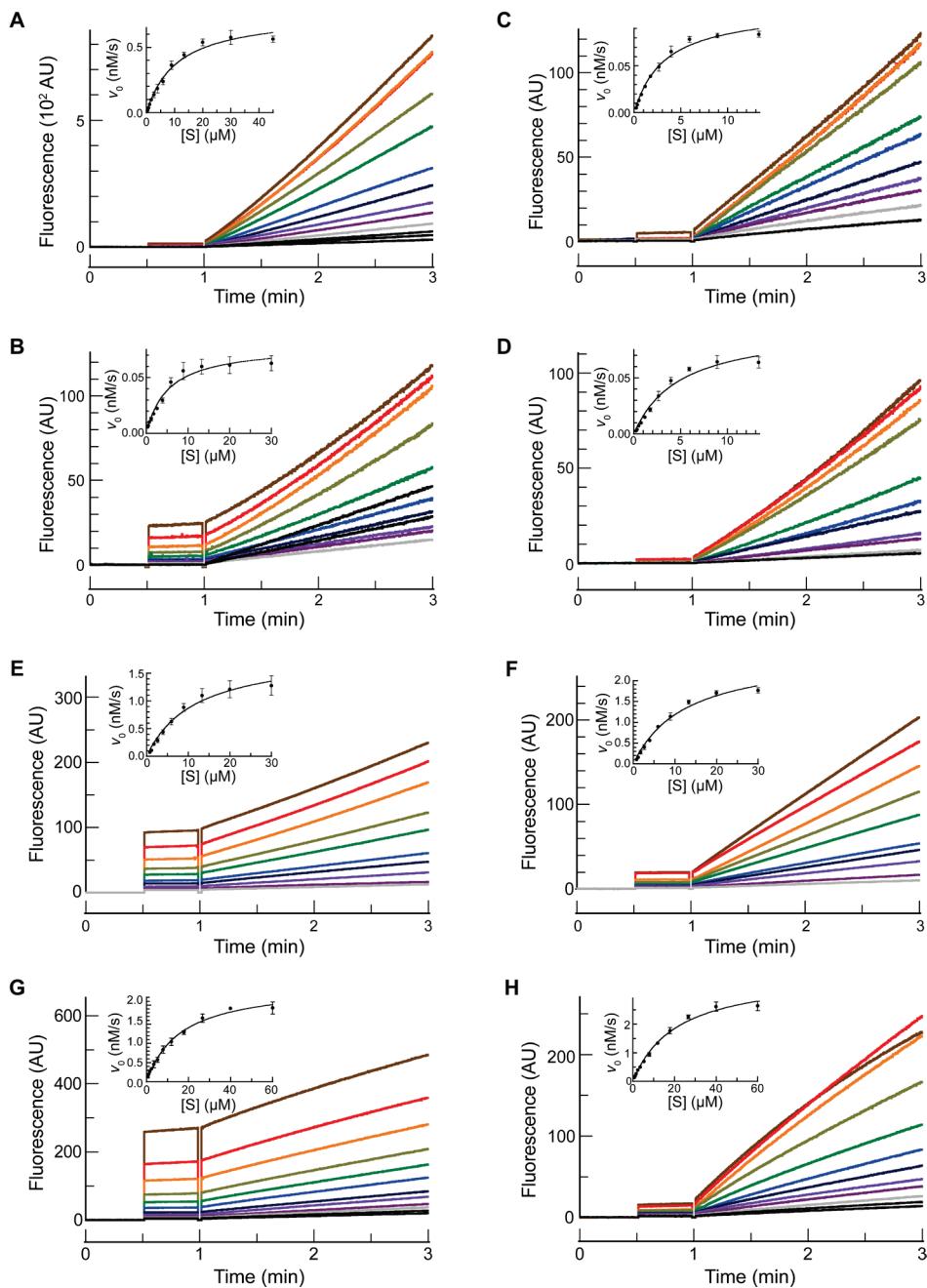


Fig. S3 Kinetic traces and Michaelis–Menten plots (inset) for the hydrolysis of profluorophores by pig liver esterase (8.3 ng/mL) in 10 mM HEPES buffer at pH 7.3. Substrate was added at 30 s; enzyme was added at 1 min. (A) Acetate ester **2a** (45 μ M \rightarrow 347 nM); $k_{cat}/K_M = 1.4 \times 10^6$ M $^{-1}$ s $^{-1}$ and $K_M = 11 \mu$ M. (B) Acetate ester **2b** (30 μ M \rightarrow 347 nM); $k_{cat}/K_M = 2.9 \times 10^5$ M $^{-1}$ s $^{-1}$ and $K_M = 5.3 \mu$ M. (C) AM ether **4a** (13.3 μ M \rightarrow 231 nM); $k_{cat}/K_M = 6.8 \times 10^5$ M $^{-1}$ s $^{-1}$ and $K_M = 3.2 \mu$ M. (D) AM ether **4b** (13.3 μ M \rightarrow 231 nM); $k_{cat}/K_M = 3.8 \times 10^5$ M $^{-1}$ s $^{-1}$ and $K_M = 4.9 \mu$ M. (E) Acetate ester **7** (30 μ M \rightarrow 780 nM); $k_{cat}/K_M = 3.2 \times 10^6$ M $^{-1}$ s $^{-1}$ and $K_M = 12 \mu$ M. (F) AM ether **6** (30 μ M \rightarrow 780 nM); $k_{cat}/K_M = 4.2 \times 10^6$ M $^{-1}$ s $^{-1}$ and $K_M = 13 \mu$ M. (G) Acetate ester **10** (60 μ M \rightarrow 694 nM); $k_{cat}/K_M = 3.1 \times 10^6$ M $^{-1}$ s $^{-1}$ and $K_M = 16 \mu$ M. (H) AM ether **9** (60 μ M \rightarrow 694 nM); $k_{cat}/K_M = 3.6 \times 10^6$ M $^{-1}$ s $^{-1}$ and $K_M = 21 \mu$ M.

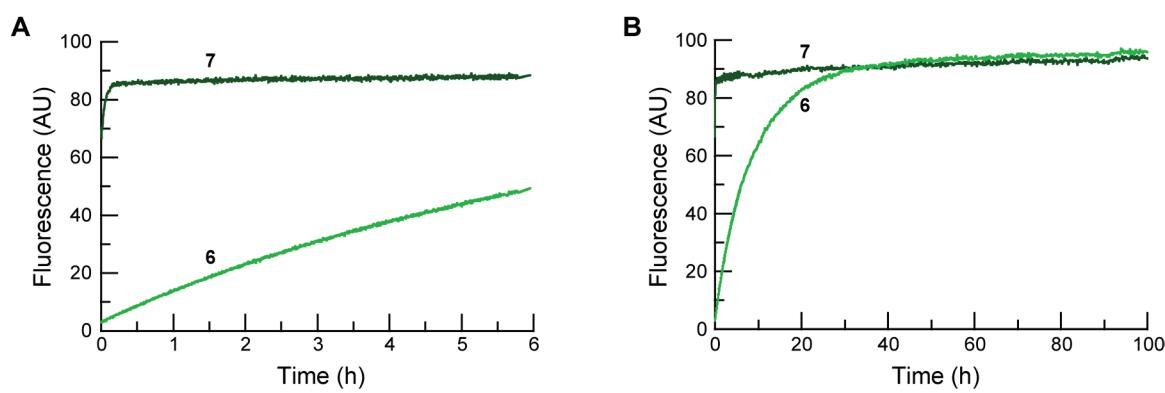


Fig. S4 Time course for the spontaneous generation of fluorescence (λ_{ex} 490 nm, λ_{em} 520 nm) from AM ether **6** and acetate ester **7** (25 nM) in DMEM containing 10% v/v FBS. (A) 0–6 h; this panel replicates Figure 3B. (B) 0–100 h.

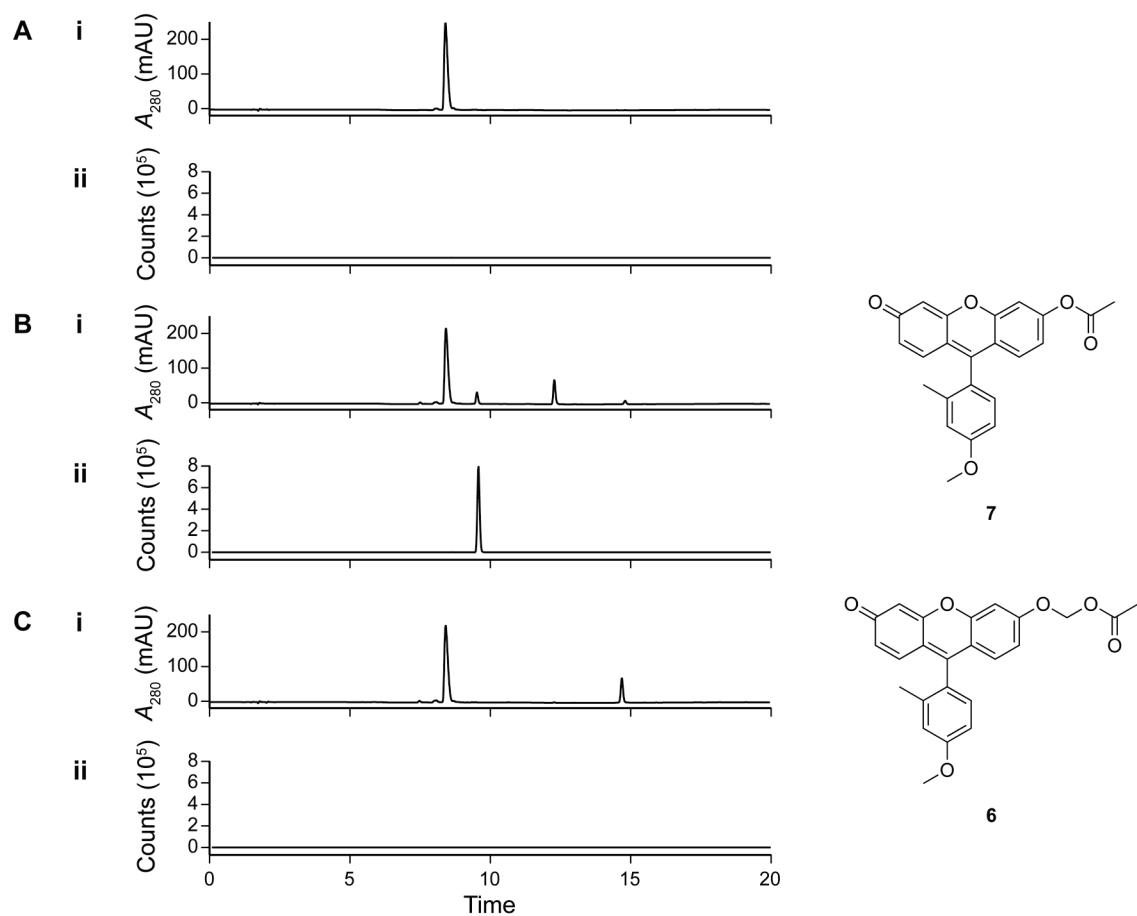


Fig. S5 Chemical reactivity of acetate ester **7** and AM ether **6** (100 μ M) with Ac-Arg-Phe-Met-Trp-Met-Lys-NH₂ (1 mg/mL) in 10 mM HEPES buffer, pH 7.3, for 2 h. (i) HPLC chromatogram of absorbance at 280 nm. (ii) LC-MS chromatogram with single ion monitoring–mass spectrometry (SIM–MS) for acylated peptide ($m/z = 981.47$). (A) Peptide only. (B) Acetate ester **7**. (C) AM ether **6**.

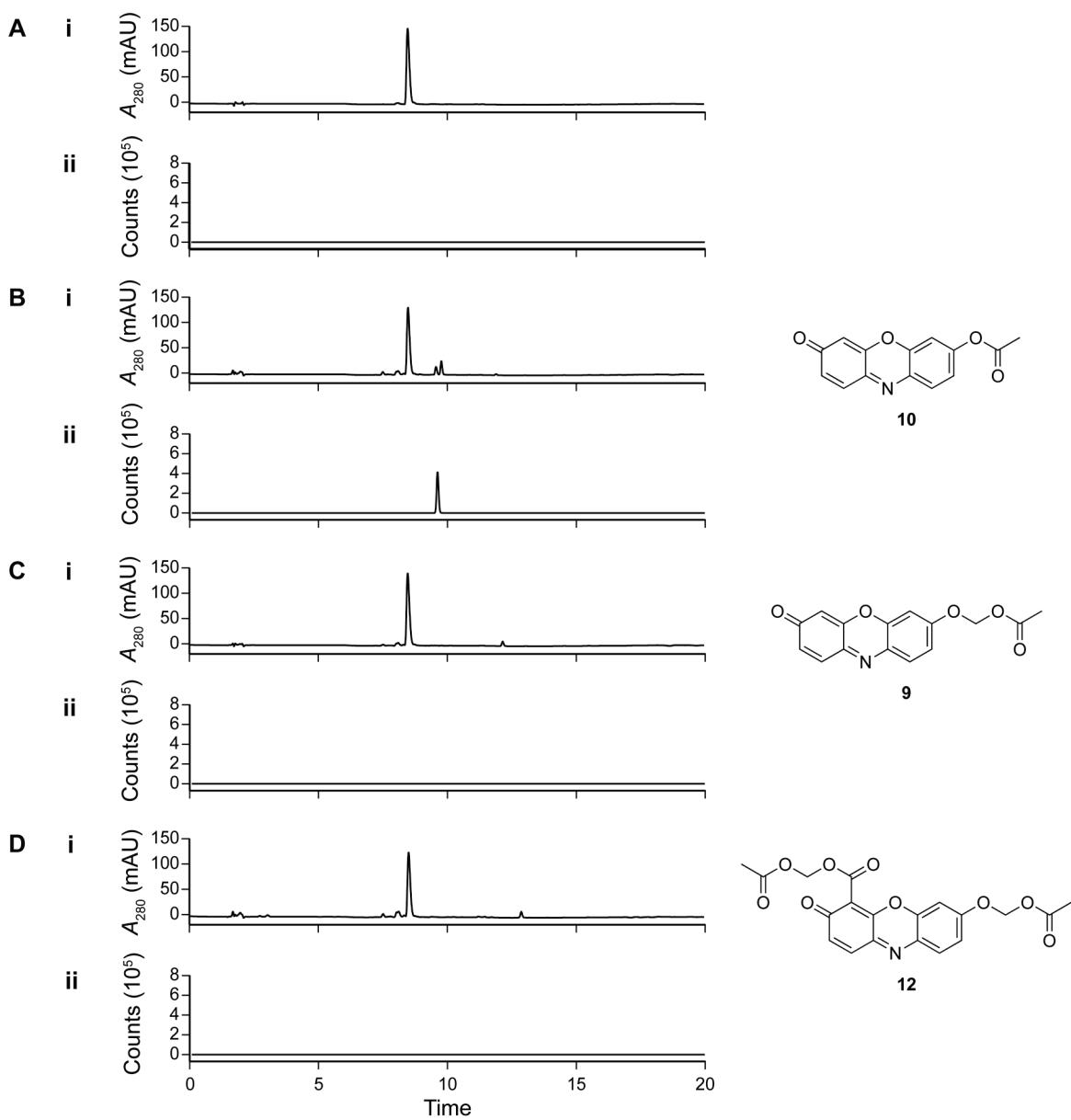
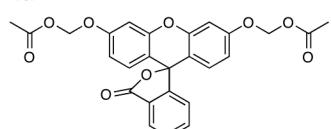
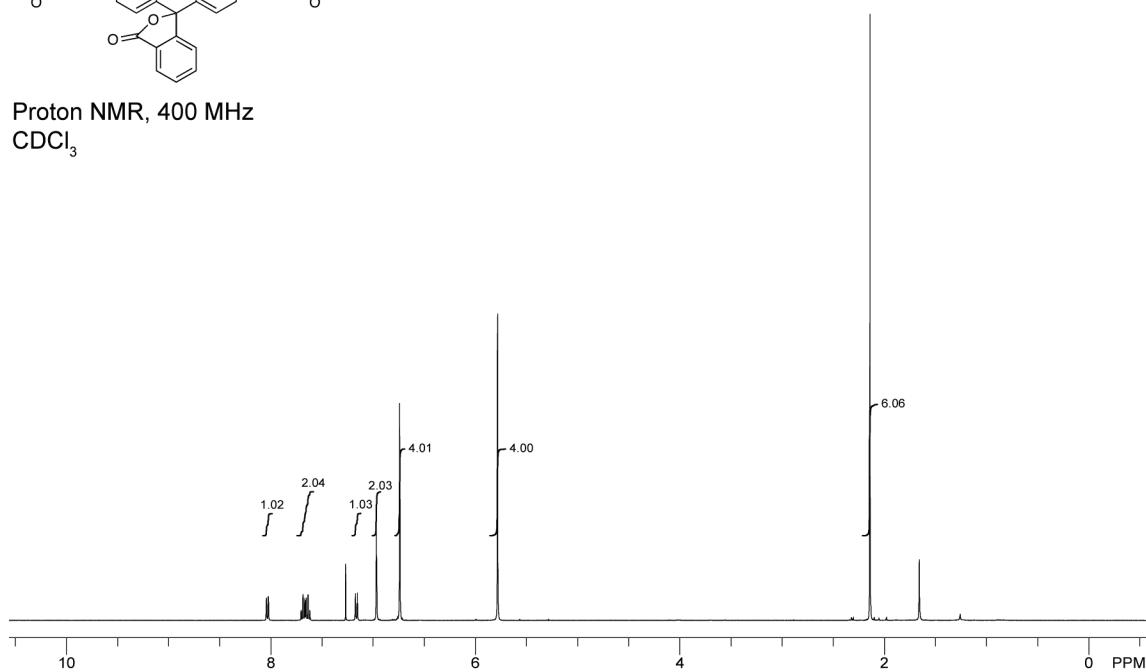


Fig. S6 Chemical reactivity of acetate ester **10** and AM ethers **9** and **12** (100 μ M) with Ac-Arg-Phe-Met-Trp-Met-Lys-NH₂ (1 mg/mL) in 10 mM HEPES buffer, pH 7.3, for 2 h. (i) HPLC chromatogram of absorbance at 280 nm. (ii) LC-MS chromatogram with single ion monitoring-mass spectrometry (SIM-MS) for acylated peptide ($m/z = 981.47$). (A) Peptide only. (B) Acetate ester **10**. (C) AM ether **9**. (D) AM ether **12**.

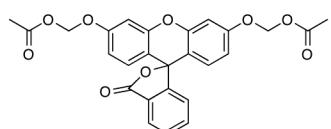
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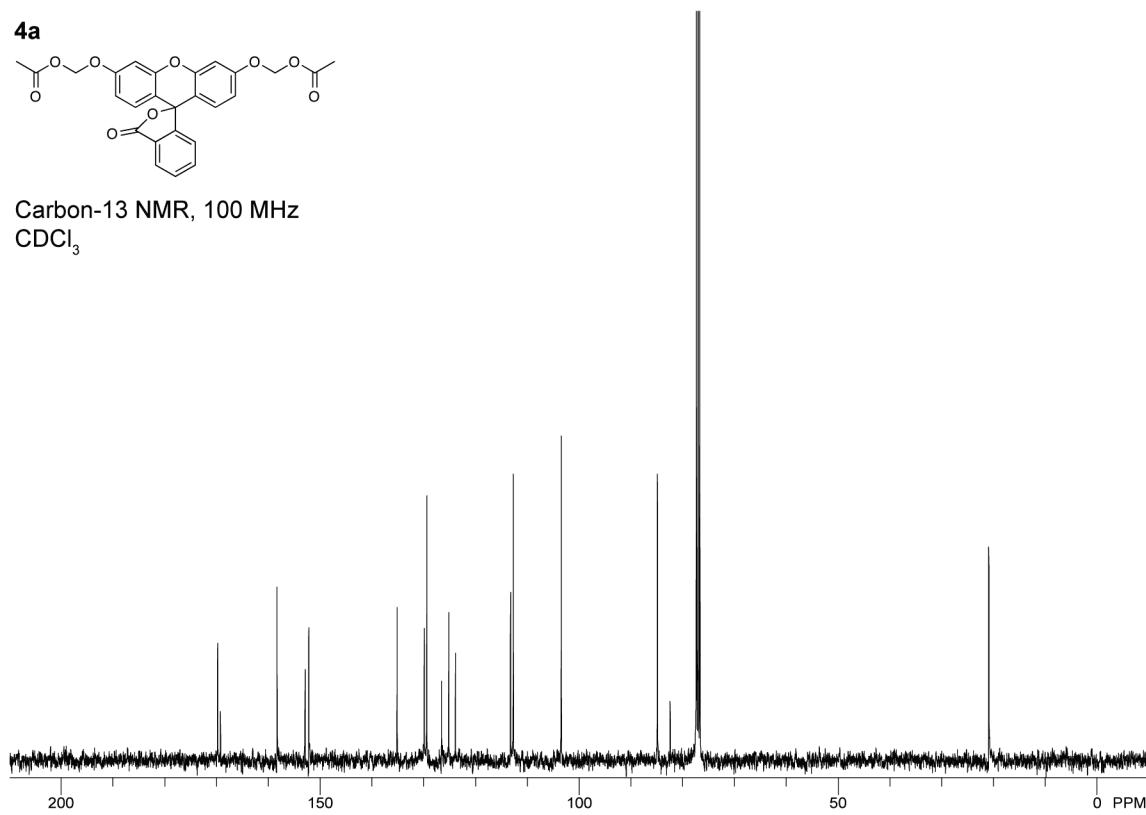
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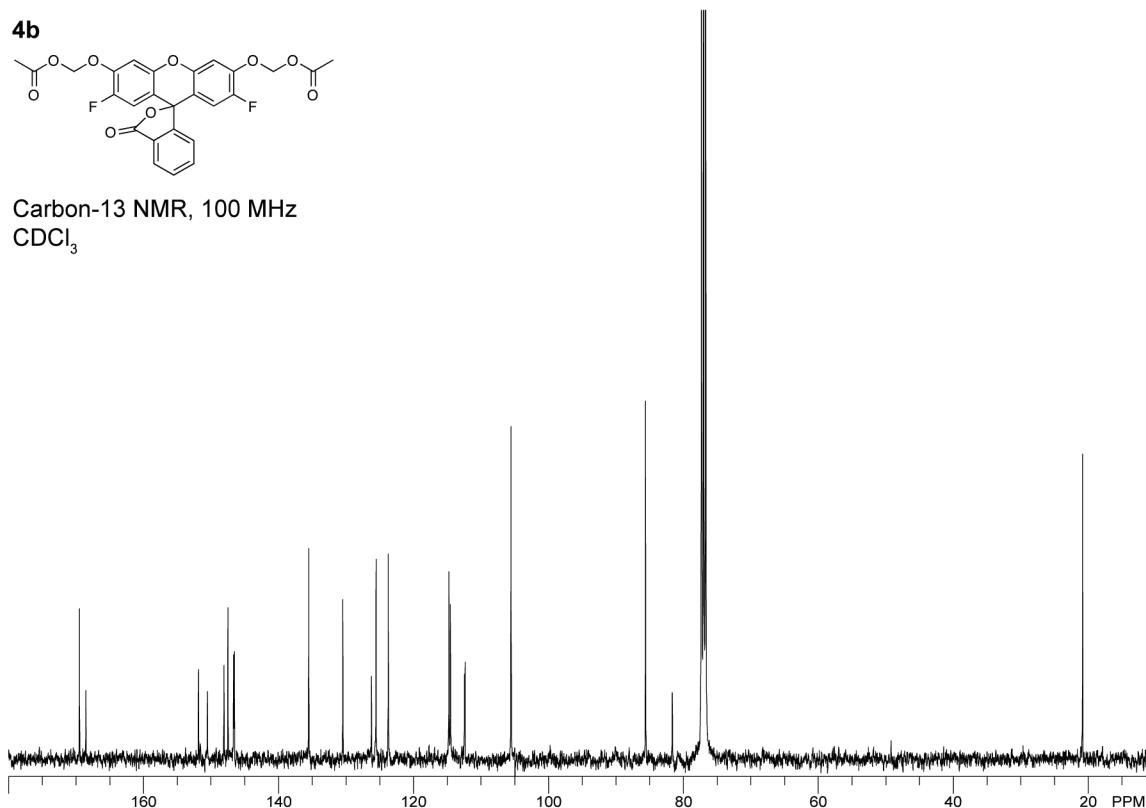
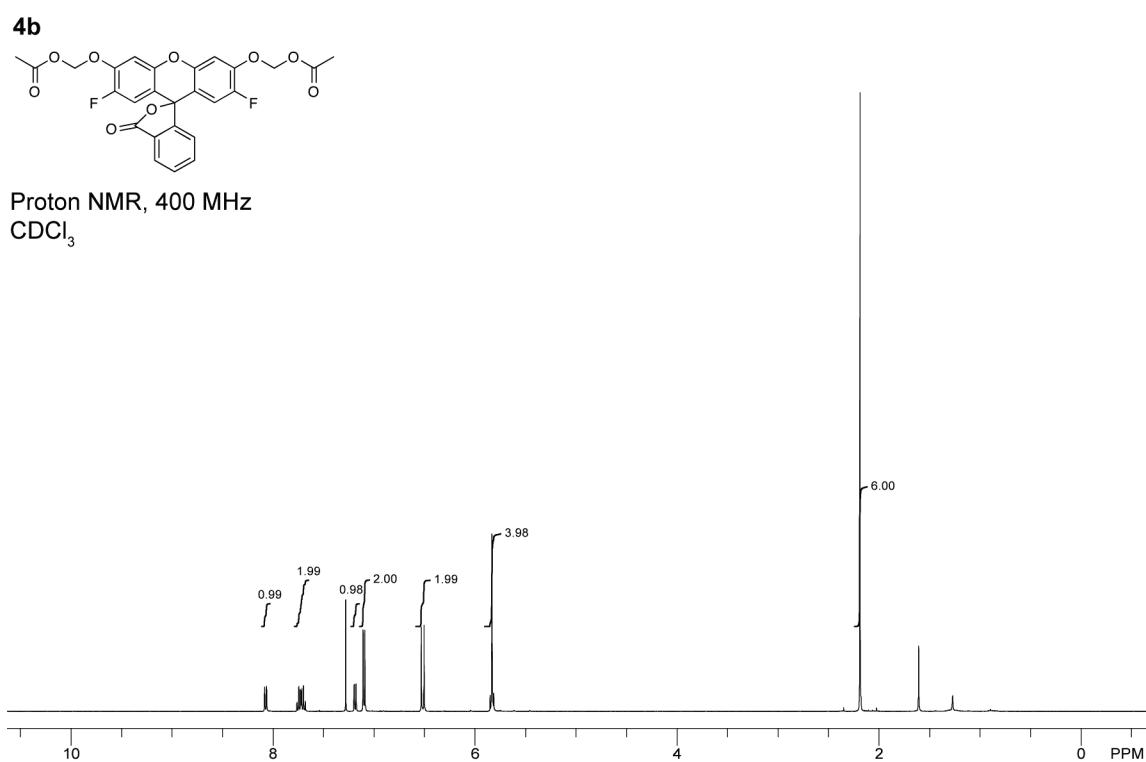


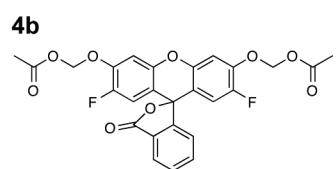
4a



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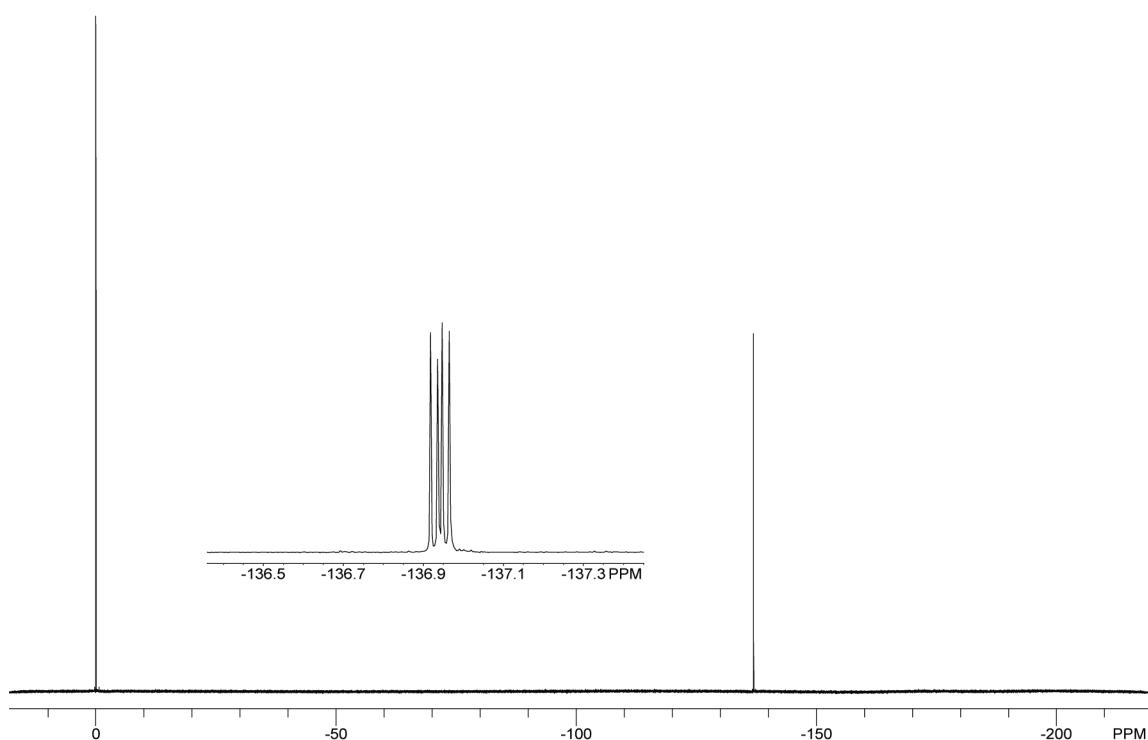


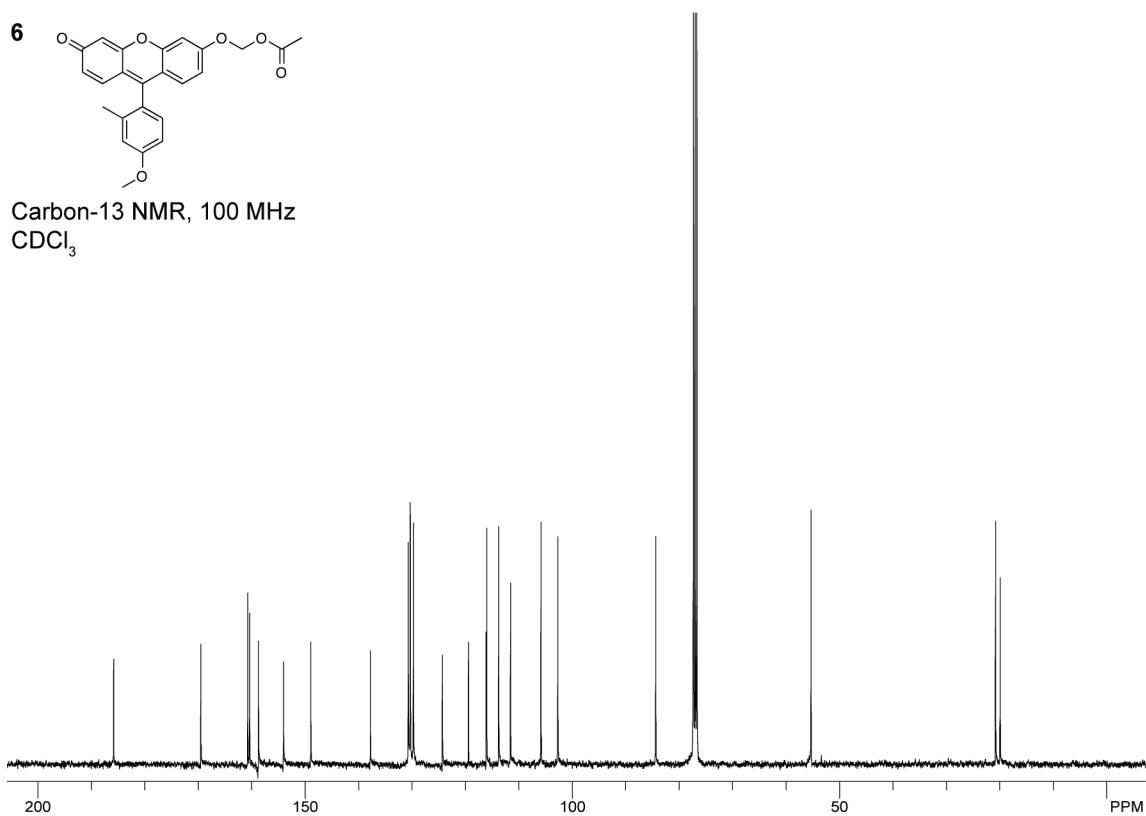
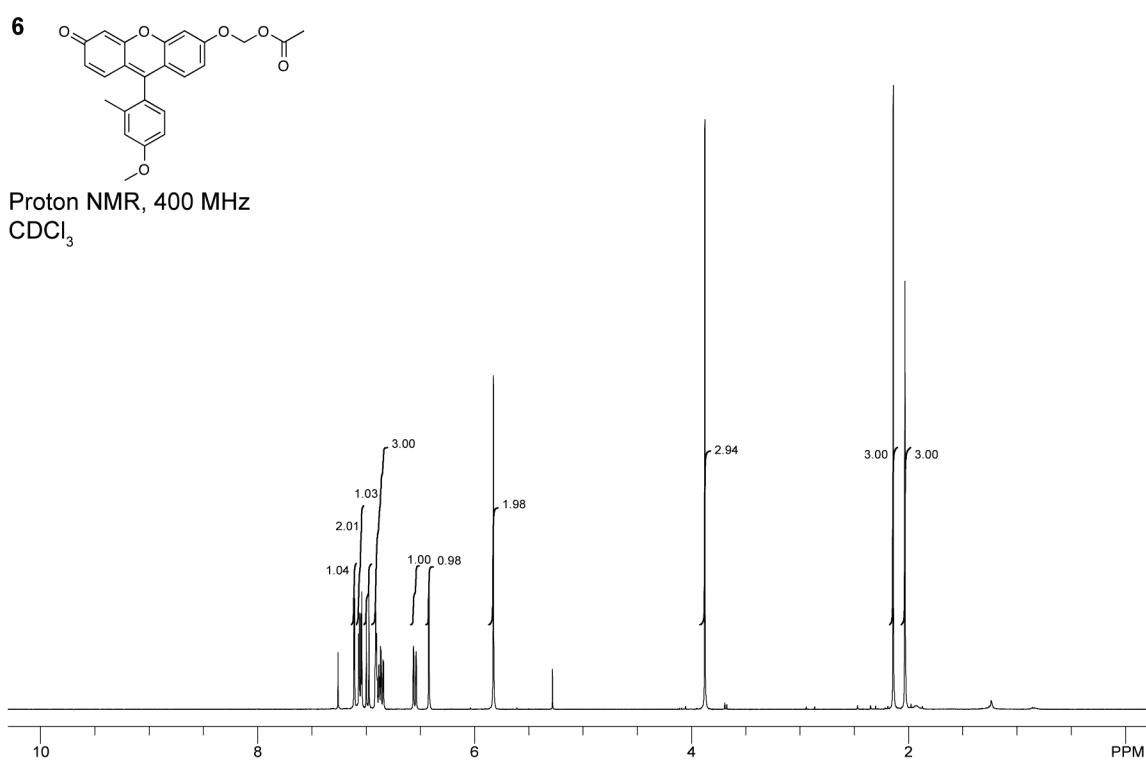


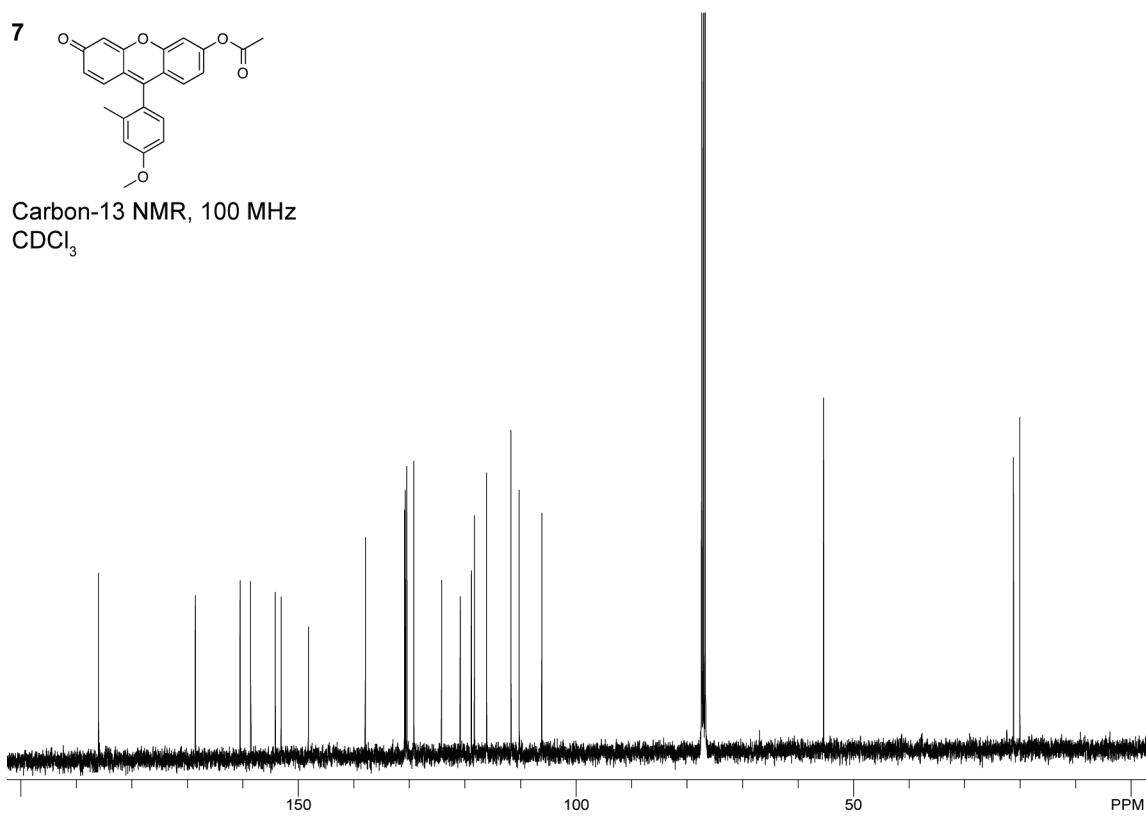
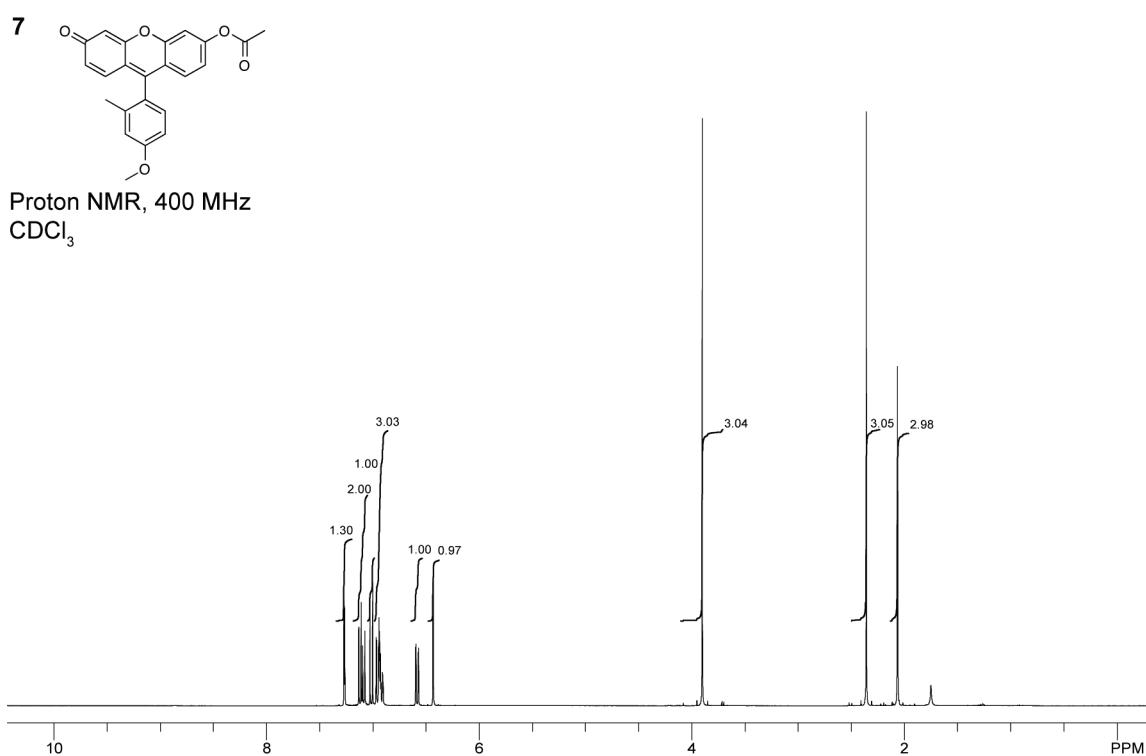


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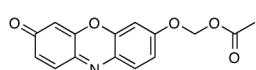
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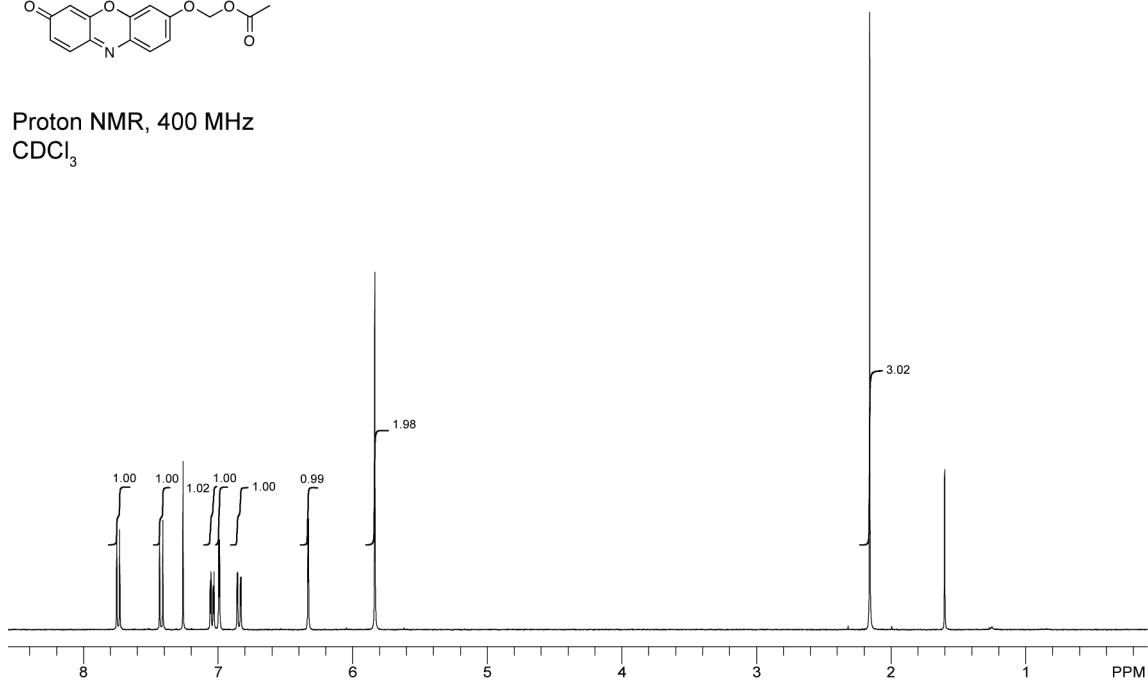




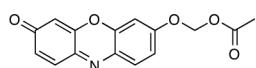
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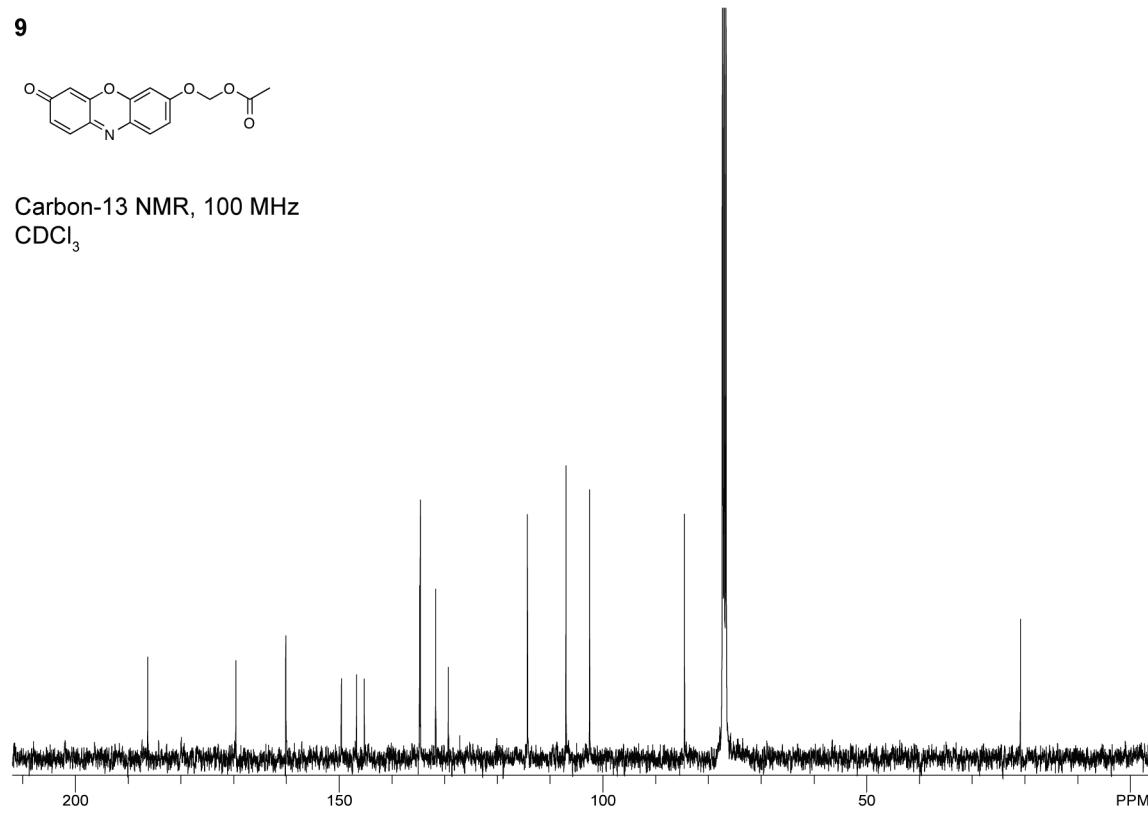
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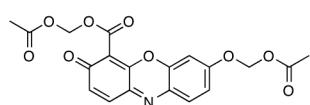
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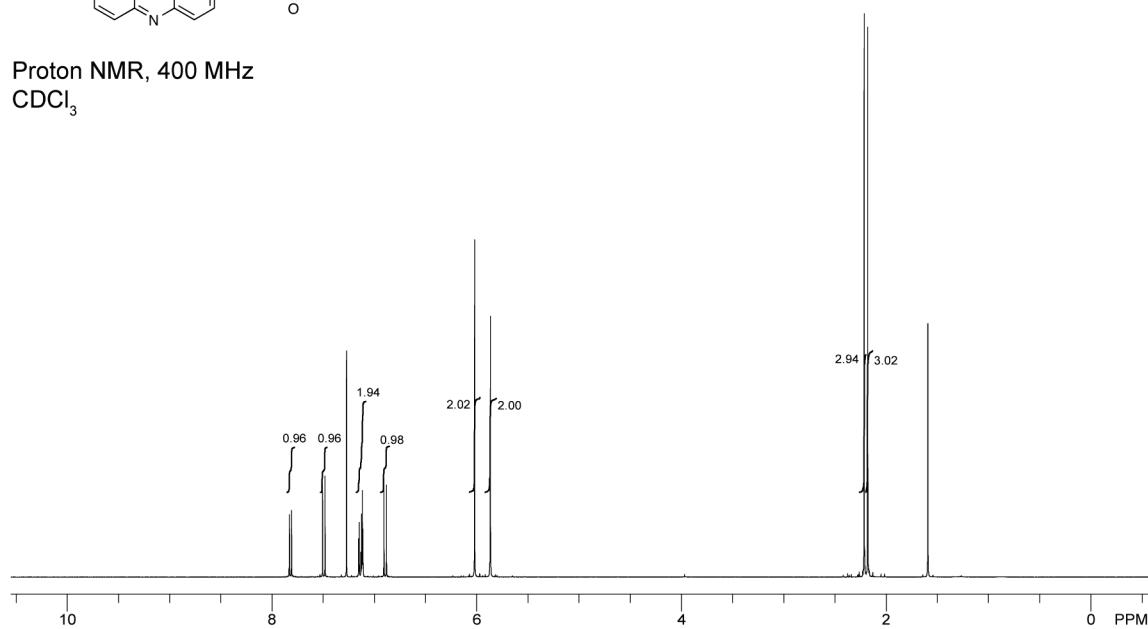
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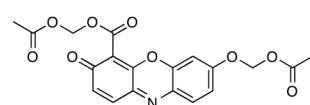
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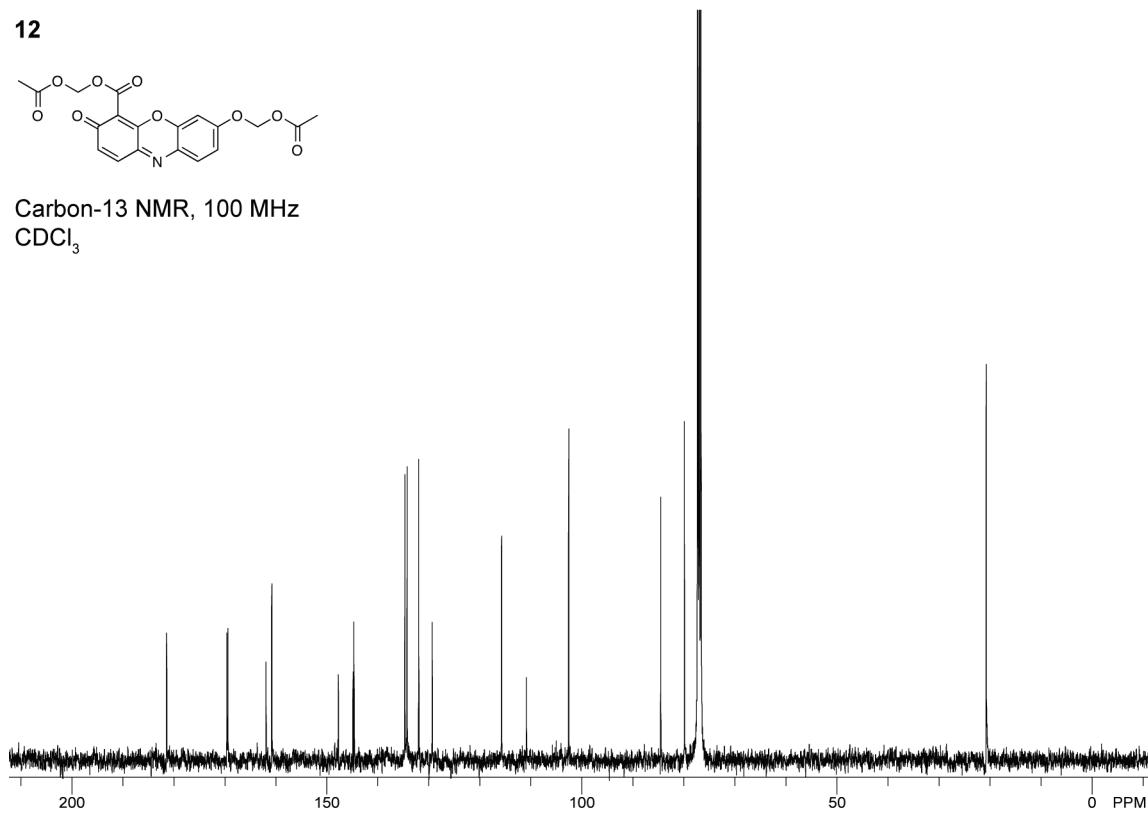
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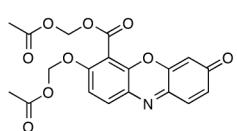
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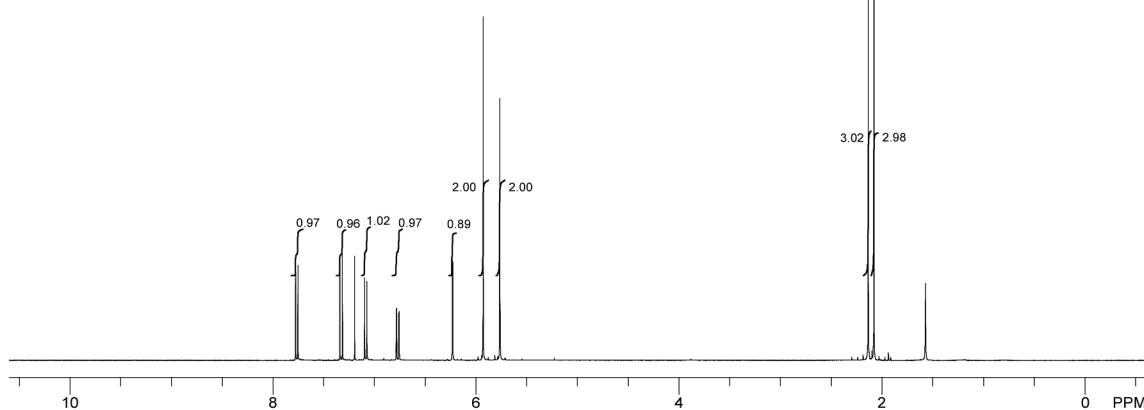
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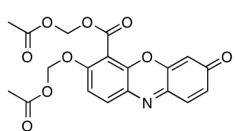
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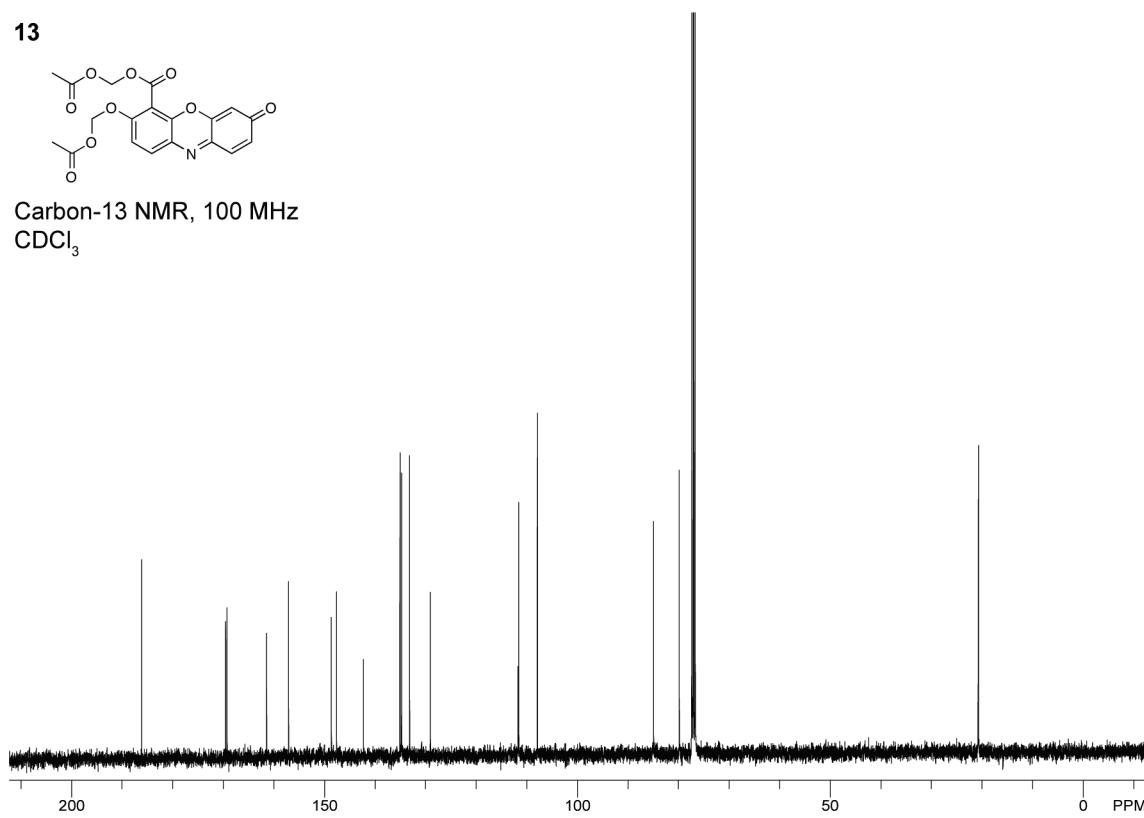
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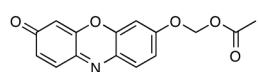
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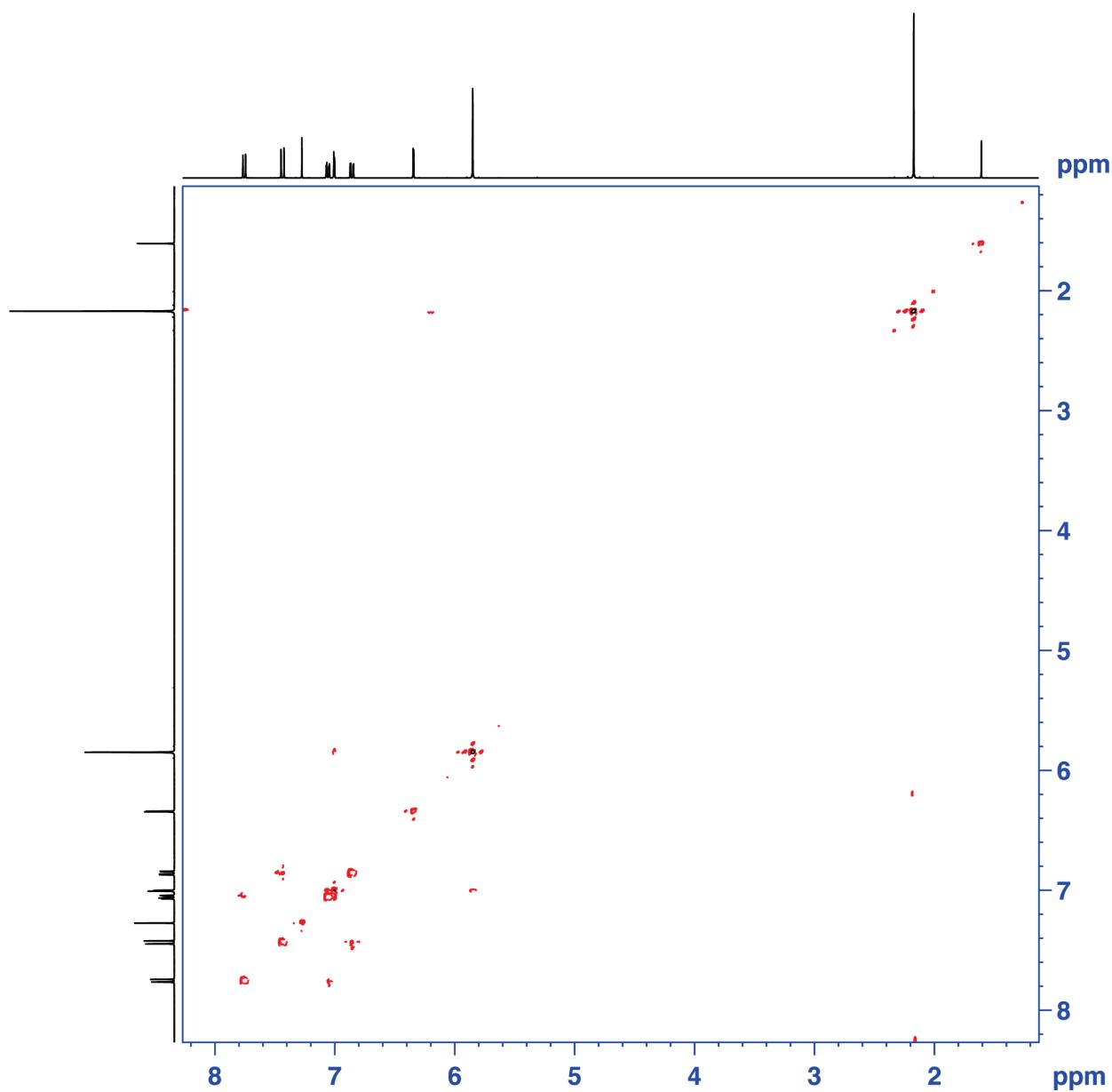
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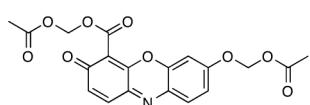
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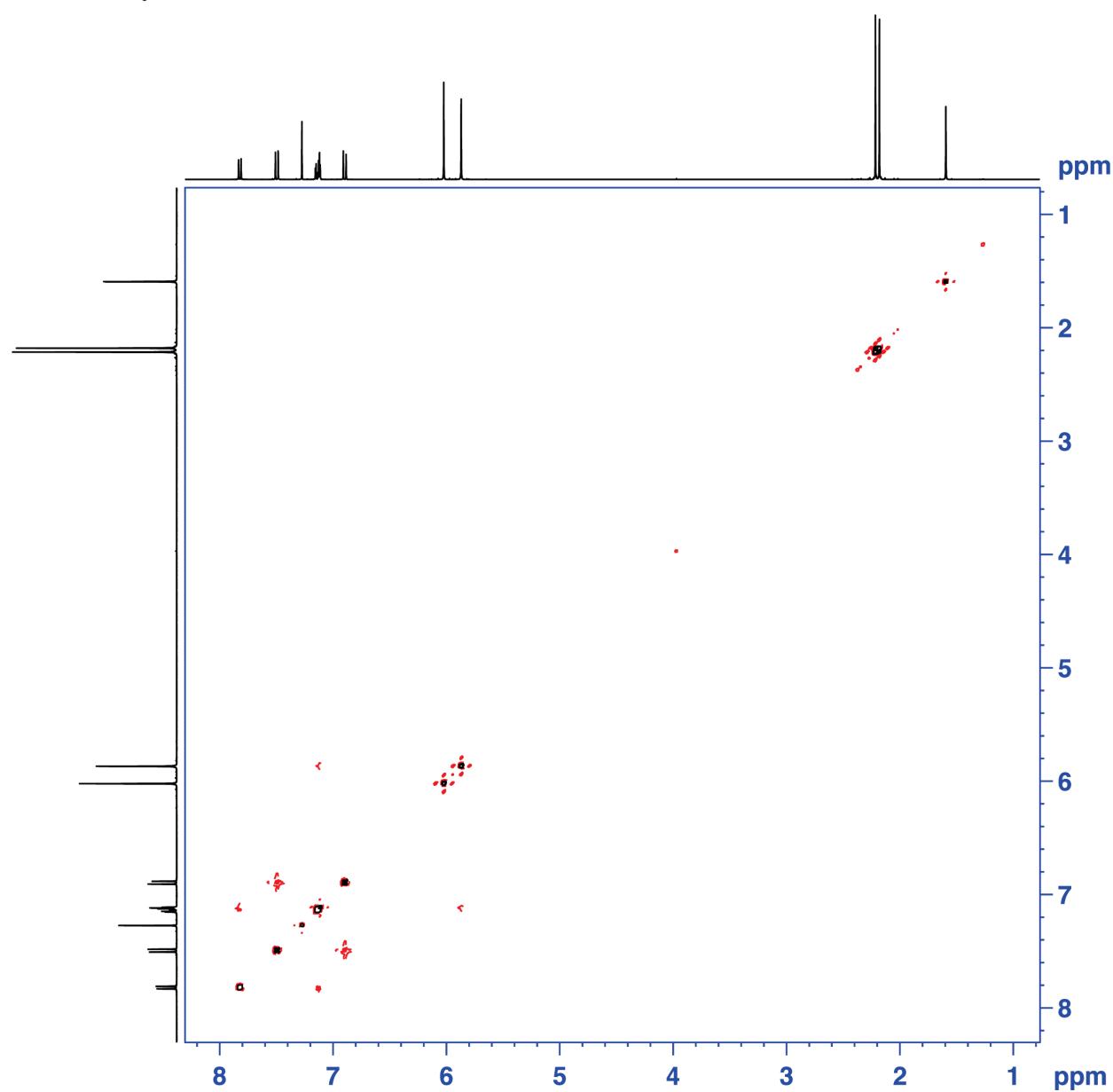
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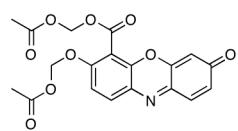
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NOESY NMR, 400 MHz
 CDCl_3



13



NOESY NMR, 400 MHz
 CDCl_3

